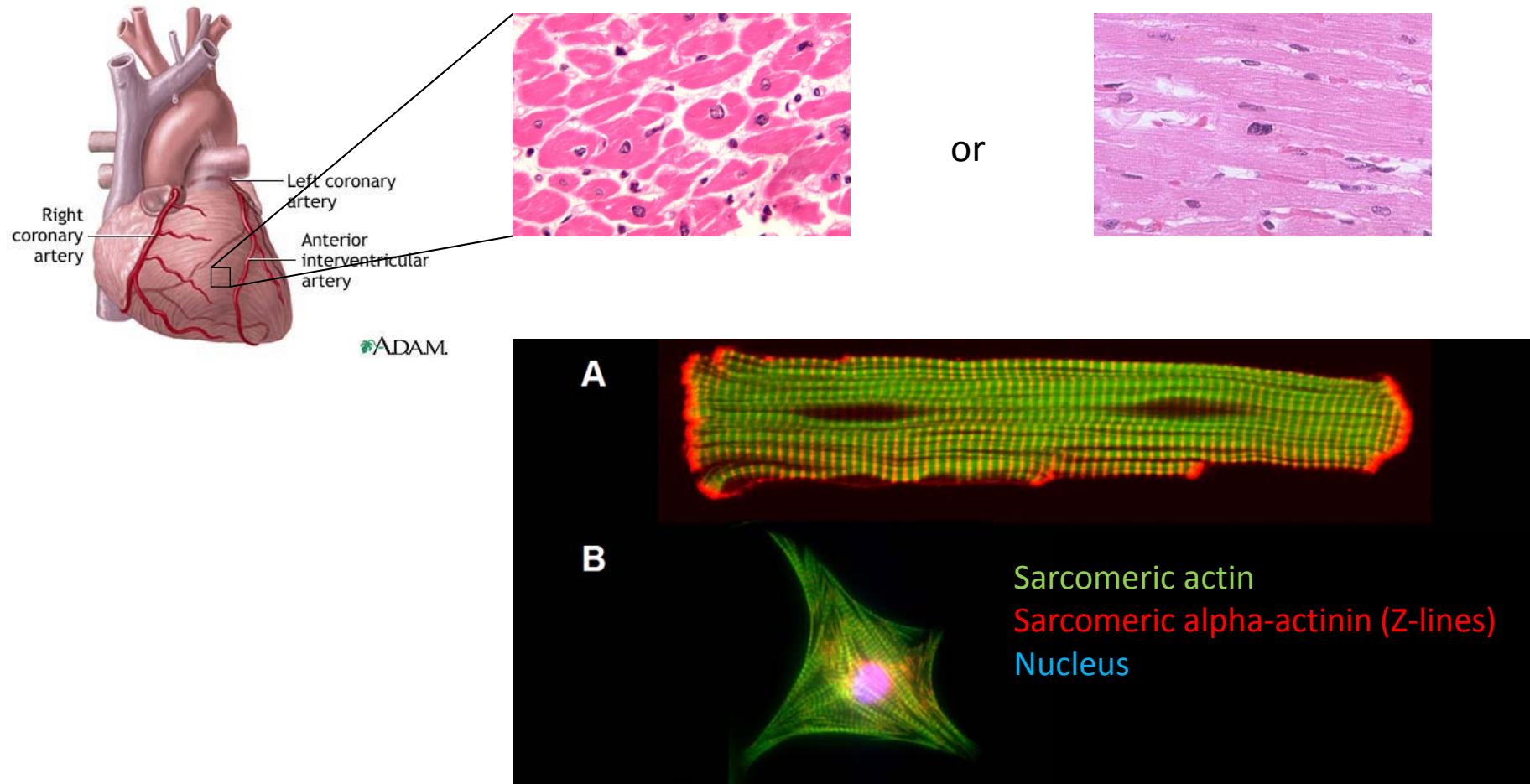


# **In vitro functional study of Cardiomyocyte**

Gwang Hyeon Eom

*Department of Pharmacology and  
Medical Research Center for Gene Regulation  
Chonnam National University Medical School,  
Gwangju, South Korea*

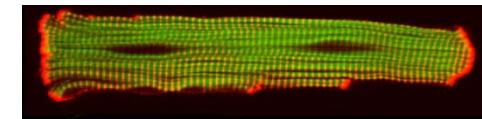
# Heart and cardiomyocyte



**A:** Adult Rat Cardiac Myocyte (nucleus not shown)  
**B:** Neonatal Rat Cardiac Myocyte

# Characteristics of cardiomyocytes

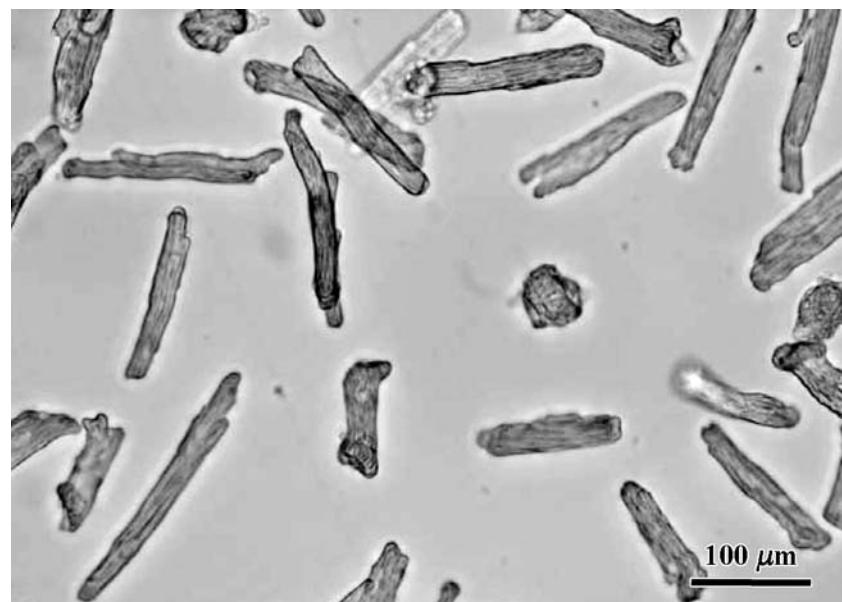
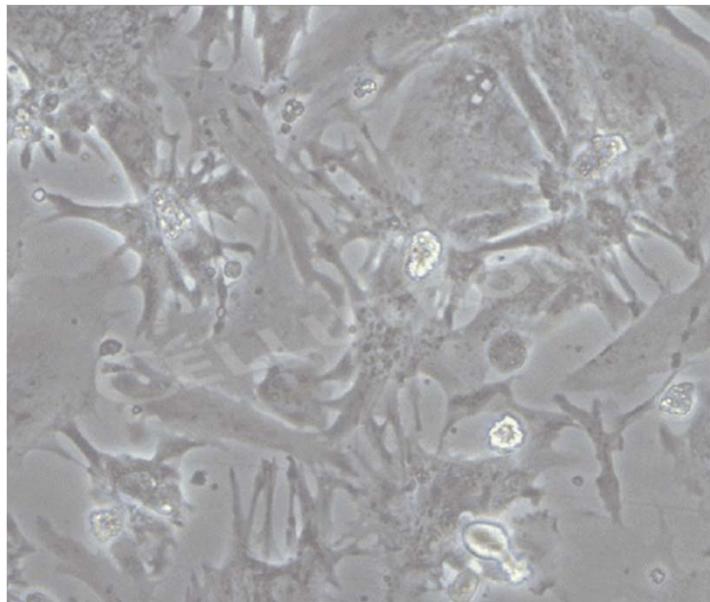
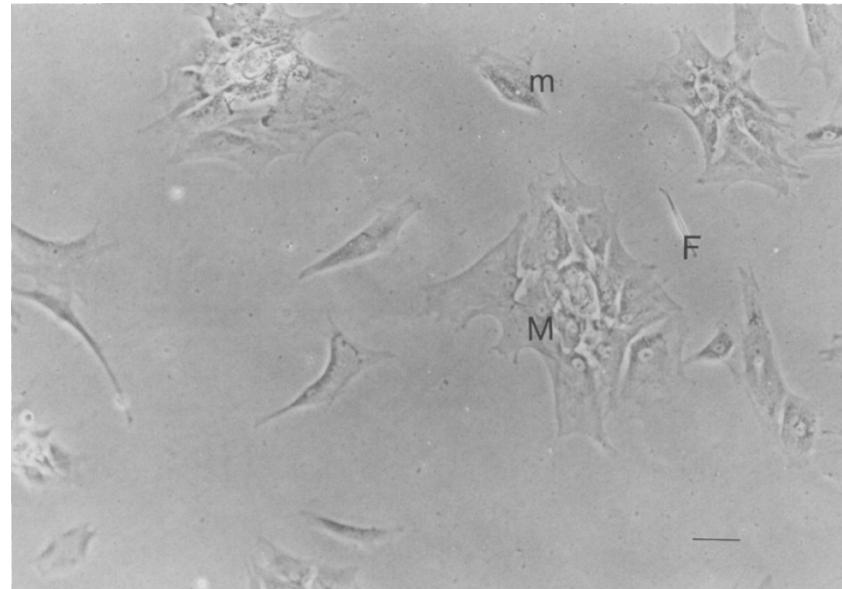
1. Involuntary striated muscle
2. Can contain one, two, or very rarely three or four cell nuclei
3. Oxygen and nutrients are supplied not by blood in the cardiac chamber directly but by coronary arteries.
4. Cardiac muscle requires extracellular calcium ions for contraction to occur
5. Until recently, it was commonly believed that cardiac muscle cells could not be regenerated. However, a study reported in the April 3, 2009 issue of Science contradicts that belief. Olaf Bergmann and his colleagues at the Karolinska Institute in Stockholm tested samples of heart muscle from people born before 1955 when nuclear bomb testing caused elevated levels of radioactive carbon 14 in the Earth's atmosphere. They found that samples from people born before 1955 did have elevated carbon 14 in their heart muscle cell DNA, indicating that the cells had divided after the person's birth.



# Sources of cardiac cells

## 1. Primary cultured

- a. Human – adult from biopsy
- b. Mouse – neonate, adult
- c. Rat – neonate, adult

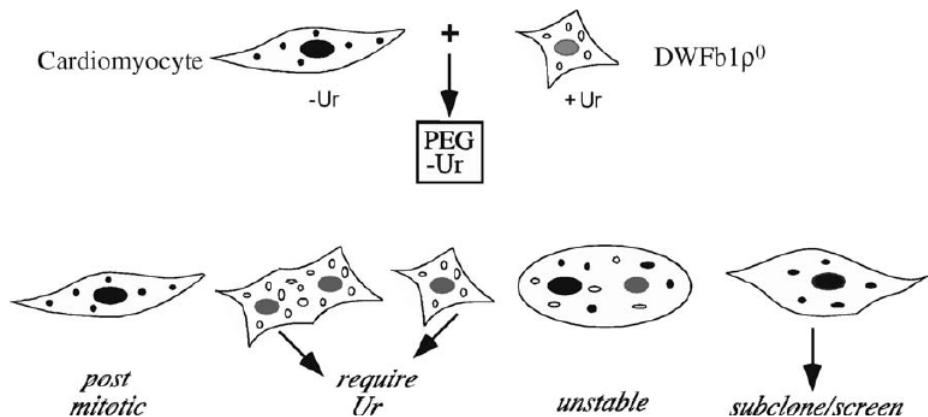


# Sources of cardiac cells

## 2. Immortalized cell line

### A. AC16 – human

- ① Adult human ventricle origin
- ② Fused with SV40 , having  
large T antigen



Comparison of AC cells and primary cardiomyocytes in culture

Markers	Primary cardiomyocytes	AC cells
<i>Culture characteristics</i>		
Cell division	NO	YES
Differentiation	NO	YES
Large T-Ag	-	+
<i>Contractile proteins</i>		
$\alpha$ -MHC	+	+
$\beta$ -MHC	+	+
VMLC-1	+	+
Troponin I	+	+
$\alpha$ -Cardiac actin	+	+
$\alpha$ -Skeletal actin	-	-
Desmin	+	+
Vimentin	-	-
$\alpha$ -Actinin	+	+
Myofilaments	+	+
Sarcomeres	+	-
<i>Membrane proteins</i>		
Desmoplakin	+	+
Intercalated disc	+	-
Gap junctions	+	+
Cx-43	+	+
Cx-40	+ (atrial)	+
<i>Transcription factors</i>		
GATA4	+	+
MYCD	+	+
NFATc4	+	+
<i>Signaling molecule</i>		
BMP2	+	+ <sup>a</sup>
<i>Ion channel</i>		
CACNa1C	+	+

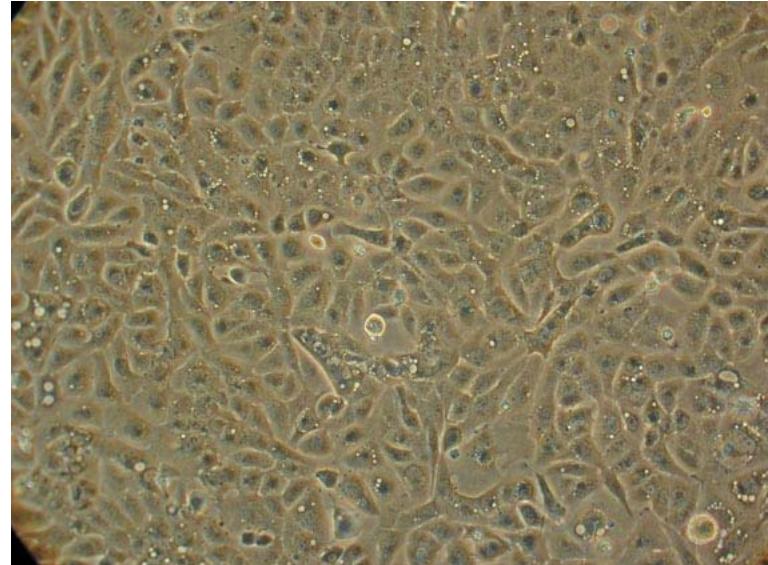
<sup>a</sup> After differentiation.

# Sources of cardiac cells

## 2. Immortalized cell line

B. HL-1 – mouse

- ① Retains contractile phenotype
- ② supplemented with
  - 2 mM L-glutamine
  - 0.1 mM norepinephrine

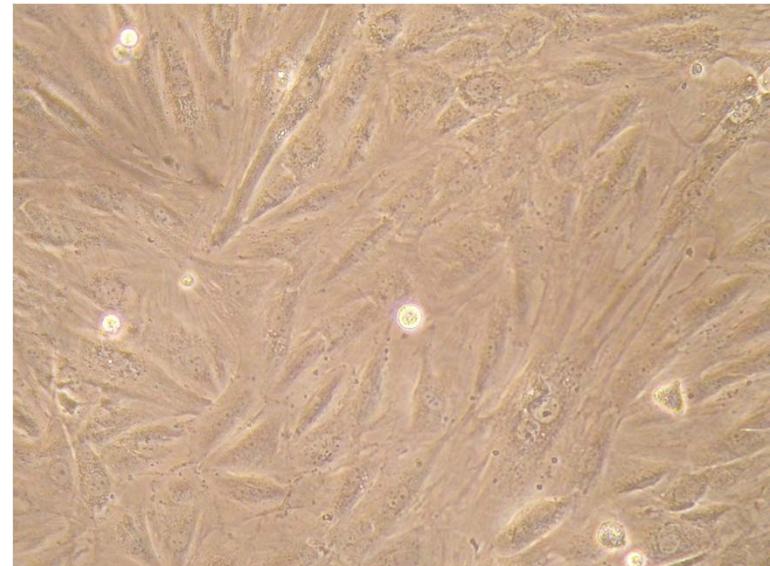


# Sources of cardiac cells

## 2. Immortalized cell line

C. H9c2 – rat

- ① Avoid to be confluent
- ② Can be differentiated to skeletal myocyte by serum deprivation or by retinoic acid



# Neonatal rat ventricular cardiomyocytes (NRVCMs)

## Materials

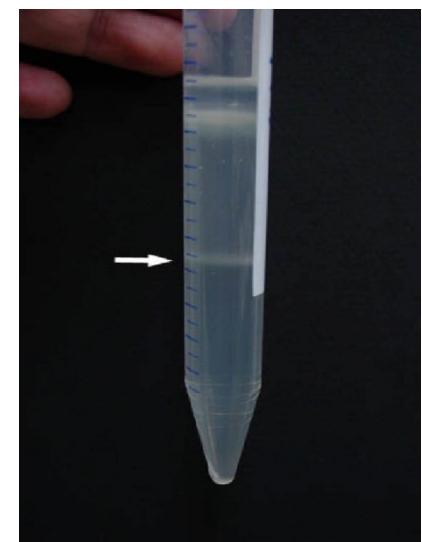
1. SD Rat pups (PN 1d ~ 2d)
2. Ads buffer – 120 mM NaCl, 6 mM glucose,  
8 mM NaH<sub>2</sub>PO<sub>4</sub>, 5 mM KCl,  
0.8 mM MgSO<sub>4</sub>, 20 mM HEPES, pH7.4
3. Collagenase (type II) – 0.5 mg/ml
4. Pancreatin – 0.6 mg/ml
5. Percoll
6. Gelatin or collagen



# Neonatal rat ventricular cardiomyocytes (NRVCMs)

## Methods

1. Isolate heart from rat pups
2. Remove appendages (use only ventricle)
3. Mince the ventricles
4. Enzyme digestion (Collagenase/Pancreatin/Ads buffer)
5. Percoll gradient  
or pre-plating without coating
6. Coat the culture plate with collagen or gelatin
7. Seed the NRVCMs  $2 \times 10^6 / \phi 60$
8. Check the NRVCMs beating

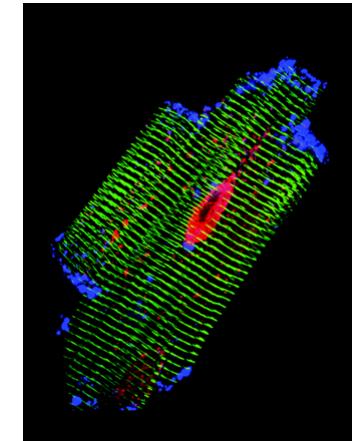


# Adult cardiac myocytes (Mouse)

## Methods

### 1. 10X Digestion buffer (DB)

NaCl	130mM
KCl	5mM
Pyruvic acid	3nM
HEPES	25mM
MgCl <sub>2</sub>	0.5mM
NaH <sub>2</sub> PO <sub>4</sub>	0.33mM
Dextrose	22mM
Adjust pH to 7.4	

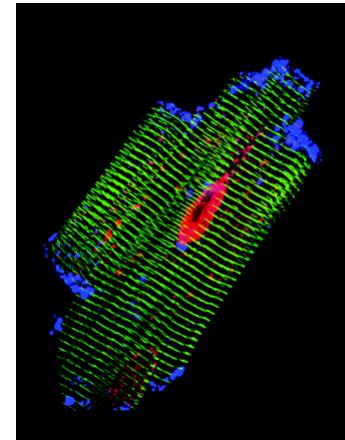


# Adult cardiac myocytes (Mouse)

## Methods

### 2. Digestion Buffer A

50mL – 1x DB with 75ul – EGTA (100mM EGTA)



### 3. Digestion Buffer B

25mL – 1x DB with 1.25uL – CaCl<sub>2</sub> (1M)

1mg – Protease, 25mg – Collagenase

### 4. Collection buffer (37°C)

2.5mL – 1X DB with 1.25ul – CaCl<sub>2</sub> (1M)

0.5mg – Protease, 15mg – Collagenase, 125mg – BSA

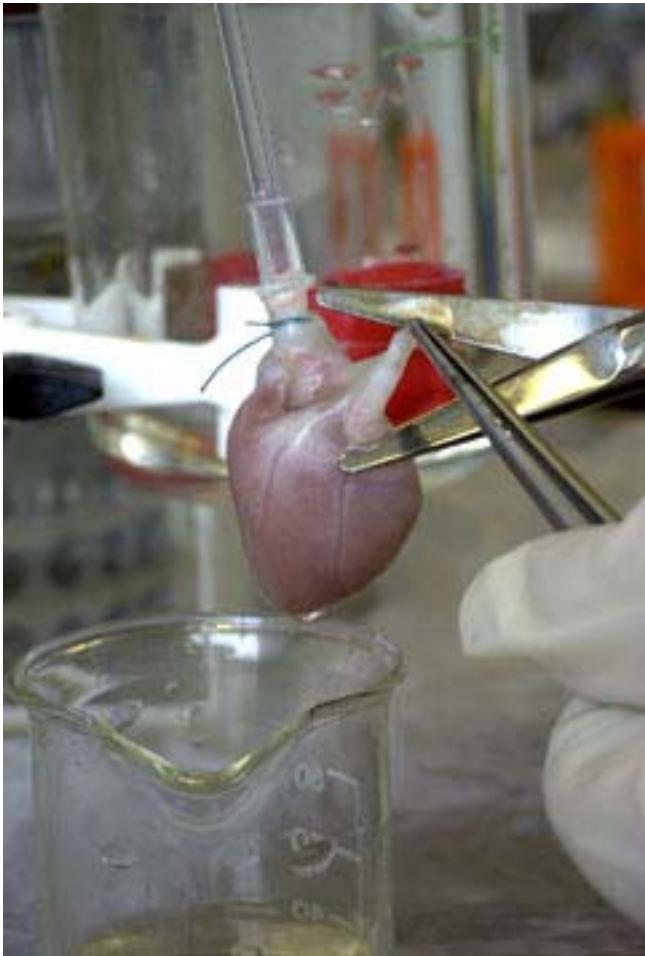
### 5. Neutralization Wash Buffer

25mL – 1X DB with 6.25ul – CaCl<sub>2</sub> (1M)

250mg BSA

# Adult cardiac myocytes (Mouse)

## Methods



ouse and keep cold with Buffer A

th ice cold Buffer A

th 37°C fluid (Buffer A) – Langendorff system

B from A after blood is washed from the heart

with scissors and place into Collection Buffer(CB)

cle into small pieces while placing into the CB

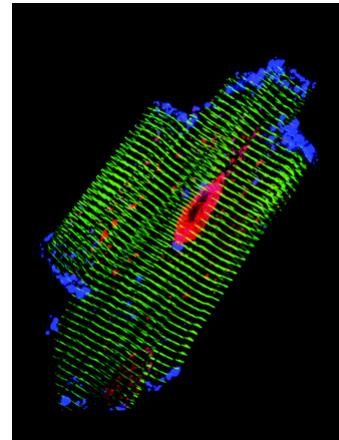
pieces sink to the bottom

e supernatant

et from the supernatant for 15 minutes at RT

atant and wash pellet by adding the

Neutralization buffer.

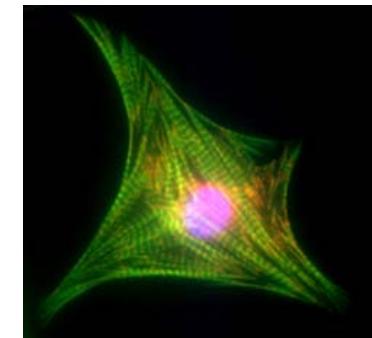


Flynn JM et al., *J Vis Exp.* 2011 Dec 28;(58):e3302. doi: 10.3791/3302.

# Limitations of NRVCMS or H9c2

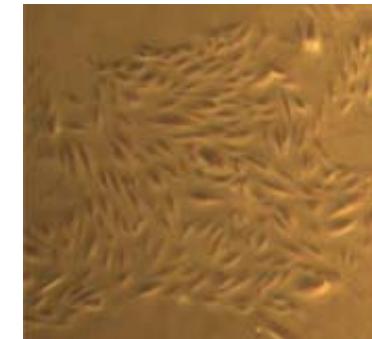
## 1. NRVCMS

- a. Risk of bacterial contamination
- b. Yield
- c. Subculture
- d. Extremely low transfection efficiency – viral delivery system



## 2. H9c2

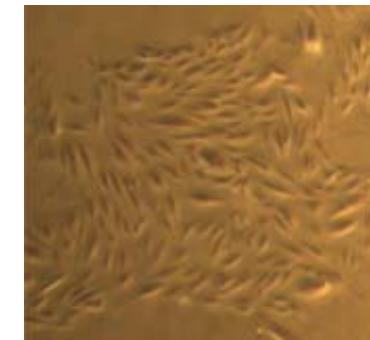
- a. Myoblast cell
- b. Lack of cardiac phenotype



# Usage of NRVCMS or H9c2

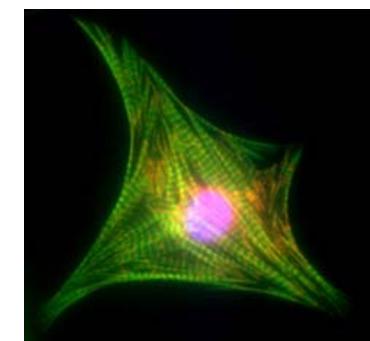
## 1. H9c2

- a. General cell works except chemical treatment



## 2. NRVCMS

- a. ICC
- b. Cell size measurement
- c. Promoter assay
- d. cell works, especially chemical study



*- when using viral delivery system, NRVCMS is the **cell of choice**.*

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## Cardiac Cells for the Advancement of Heart Research

The cardiac system provides the mechanical means for delivering nutrients to the tissues and organs of the body. Study of heart cells is key to identifying pathologies that injure or kill millions of people every year. Cell Applications, Inc. provides cardiac cells of various lineages for the advancement of heart research.

Examples of Research with Cell Applications, Inc.'s Cardiac Cells:

- Human Aortic Endothelial Cells (HAOEC) have been used as an *in vitro* model for the study of morphological and ultra-structural changes in the development of atherosclerosis.
- Human Aortic Smooth Muscle Cells (HAOSMC) are capable of synthesizing collagen, elastin, myosin and glycosaminoglycan.
- Human Cardiac Fibroblasts (HCF) has been used to study the effects of TNF-alpha and angiotensin (Ang) II levels on fibroblast function.
- Human Coronary Artery Endothelial Cells (HCAEC) are crucially involved in the regulation of coronary blood

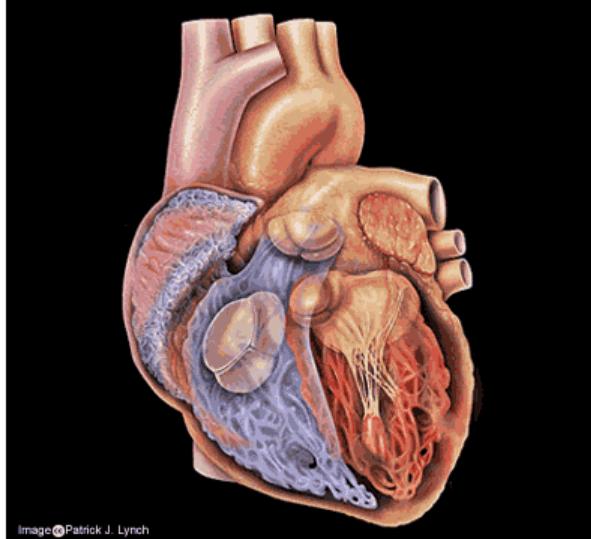


Image © Patrick J. Lynch

<http://www>

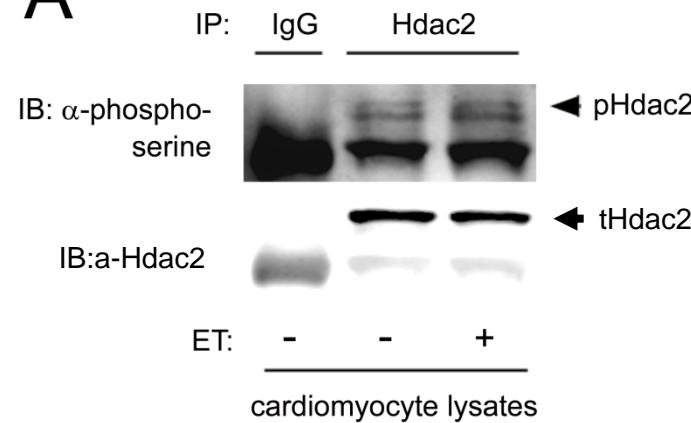
»Trader, Inc)

<http://www.cellapplications.com/primary-heart-cells> (Abba BioScience)

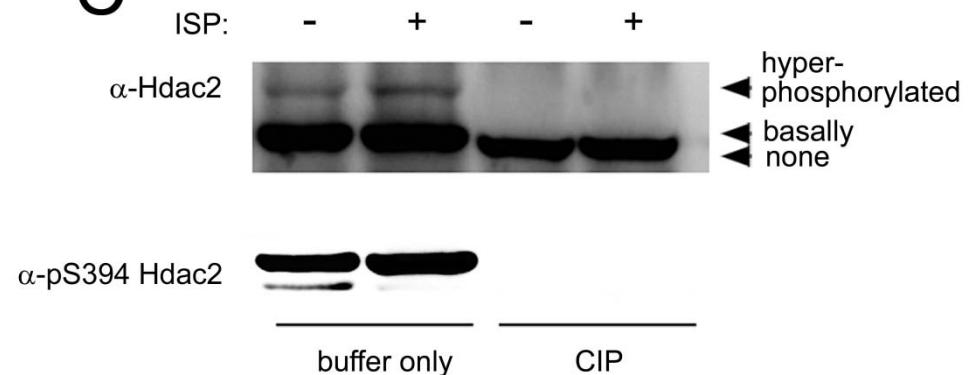
# Examples



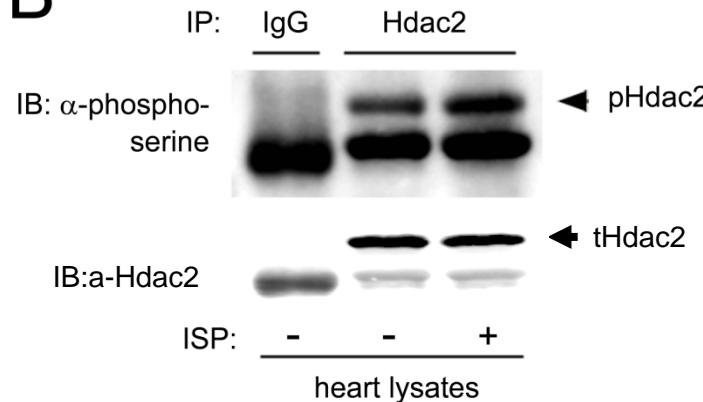
**A**



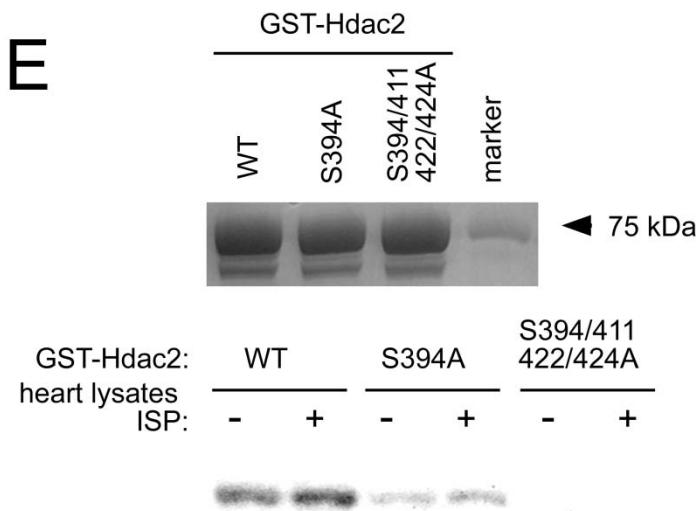
**C**

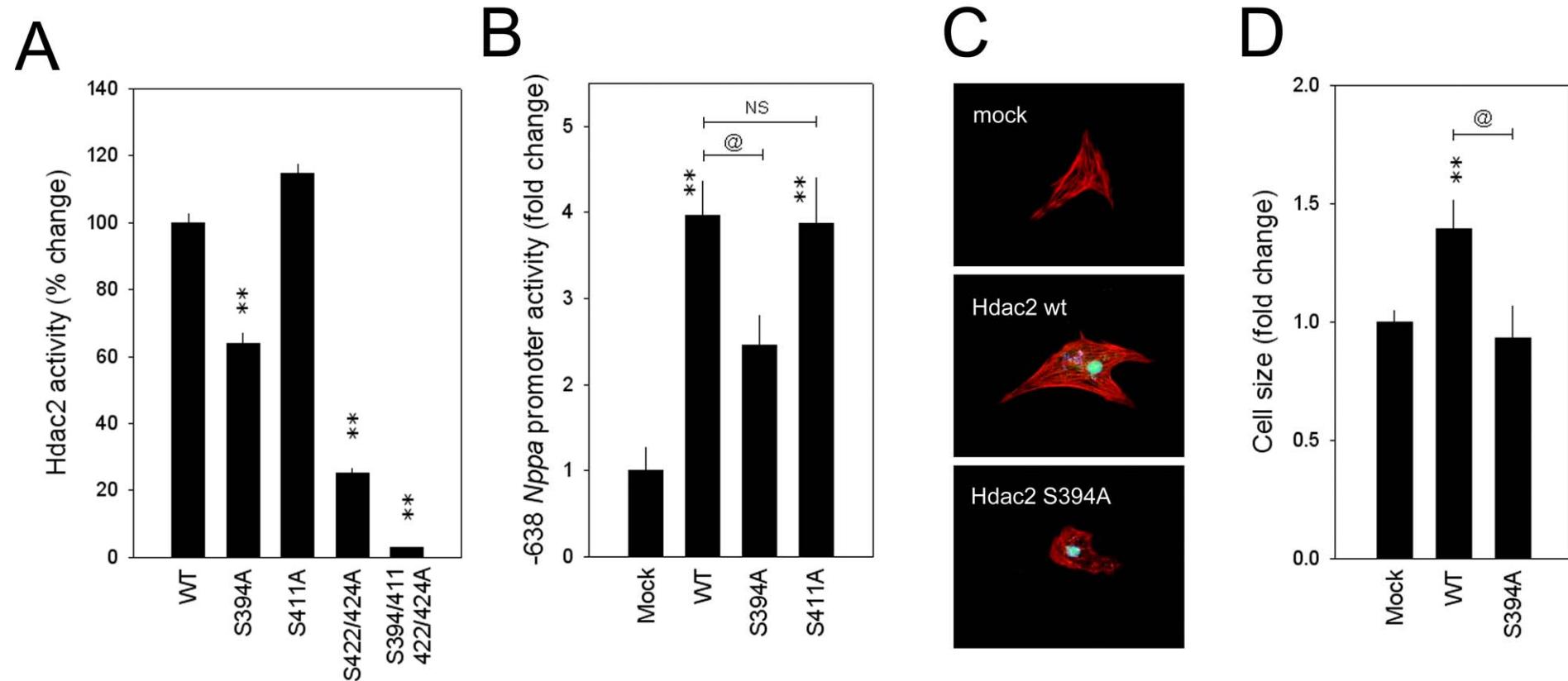
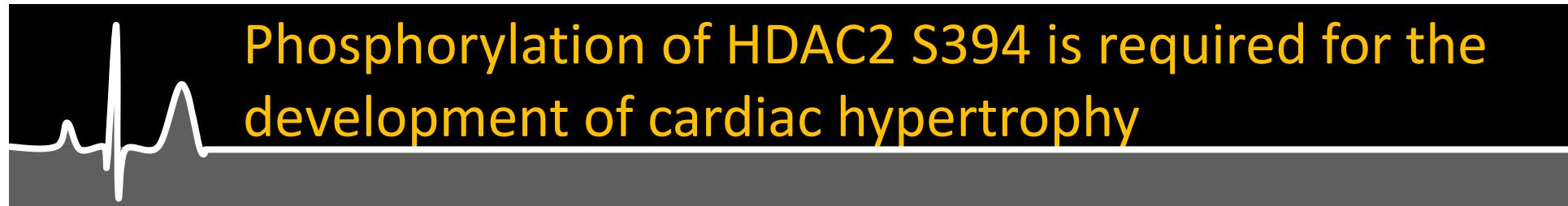


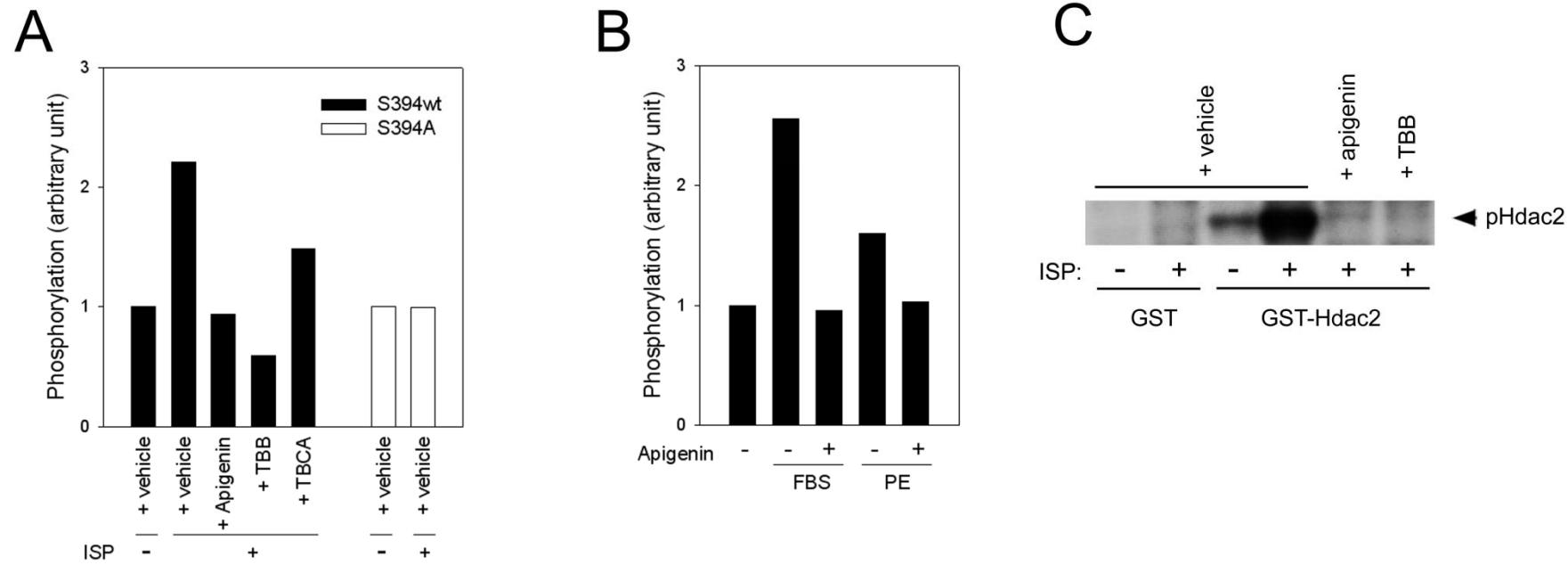
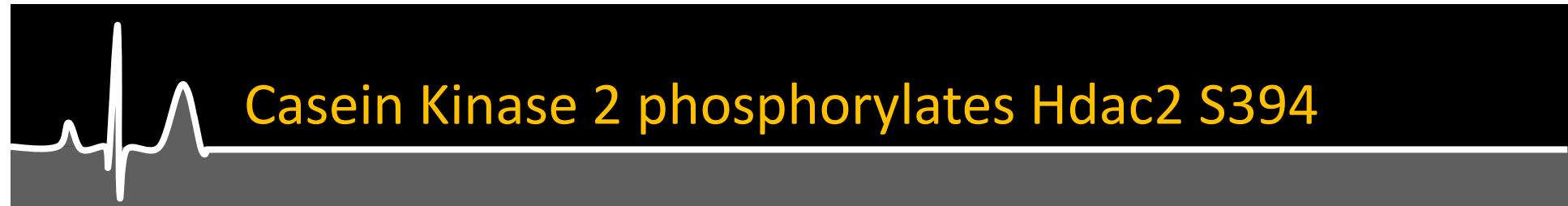
**B**



**E**

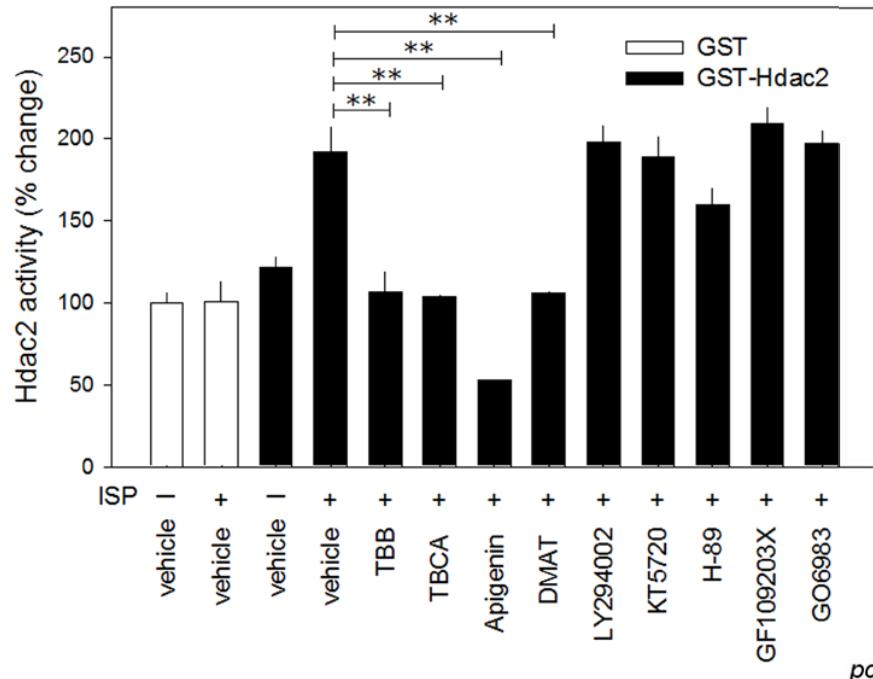




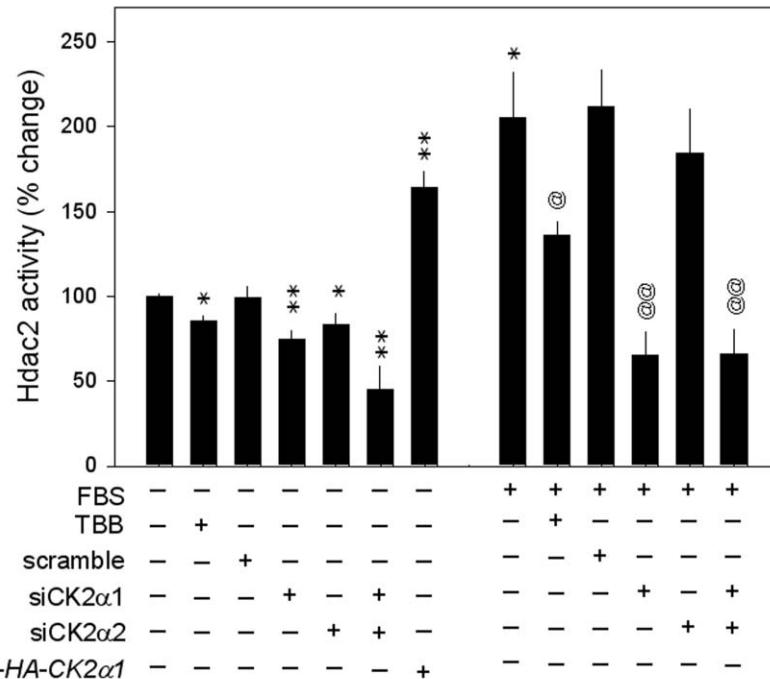


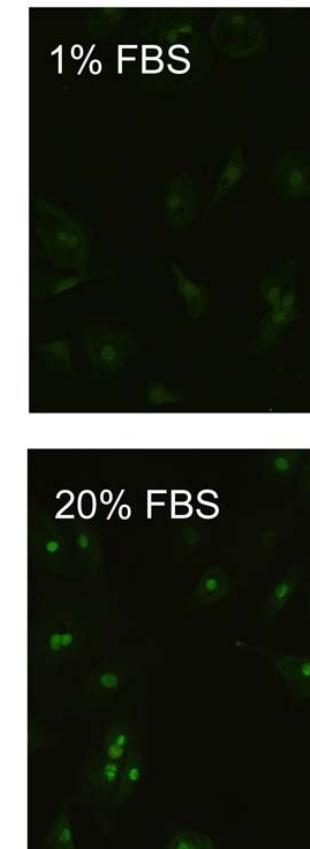
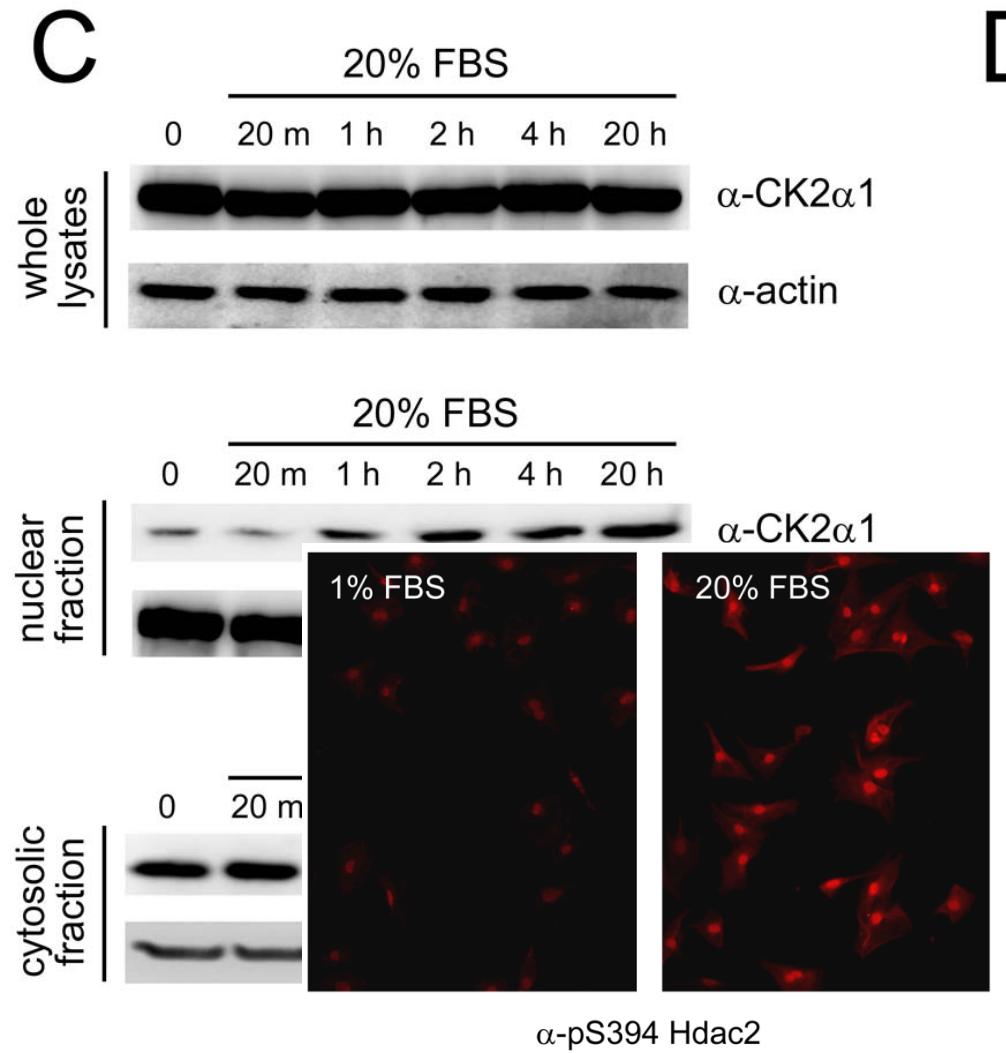


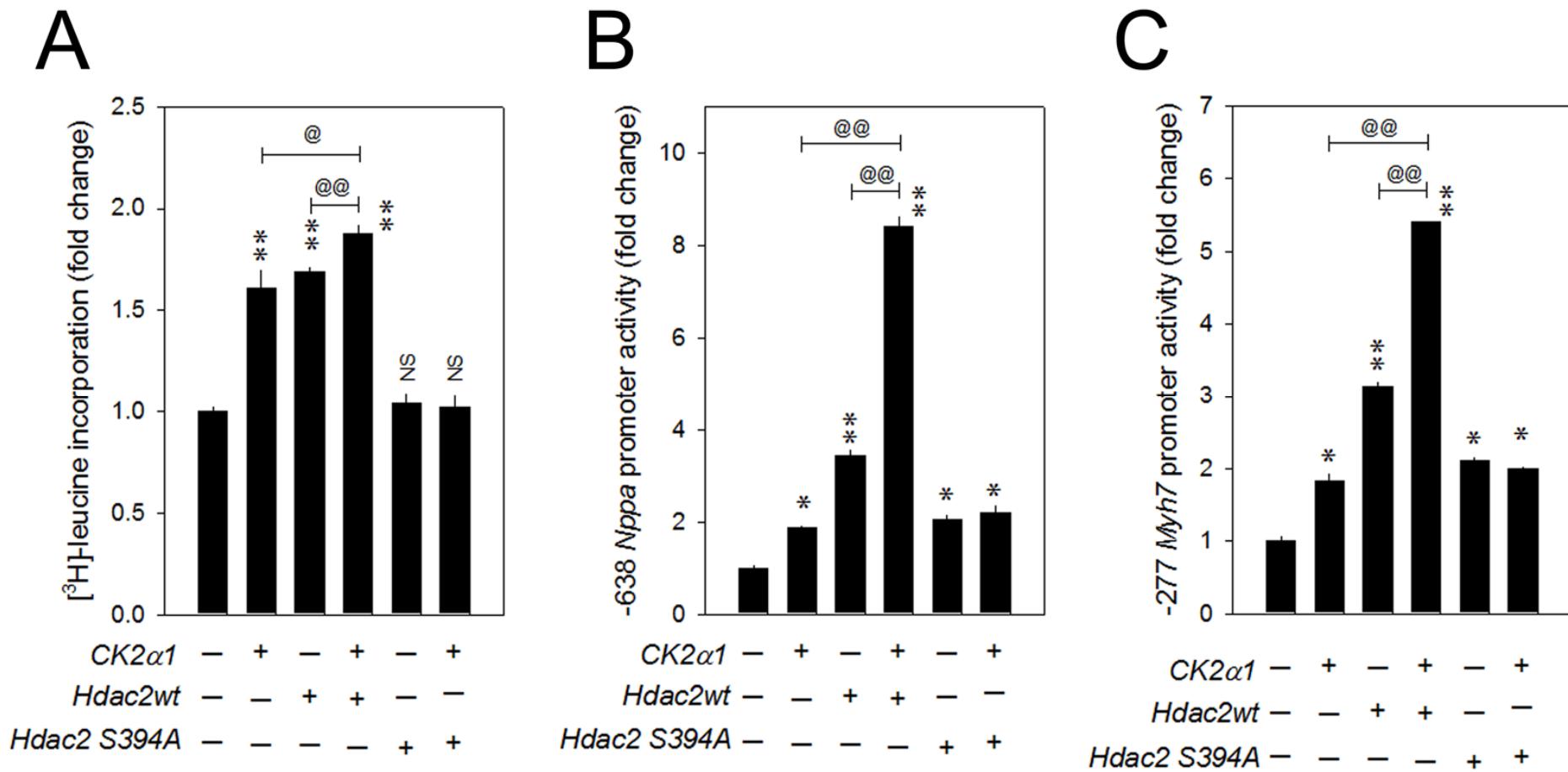
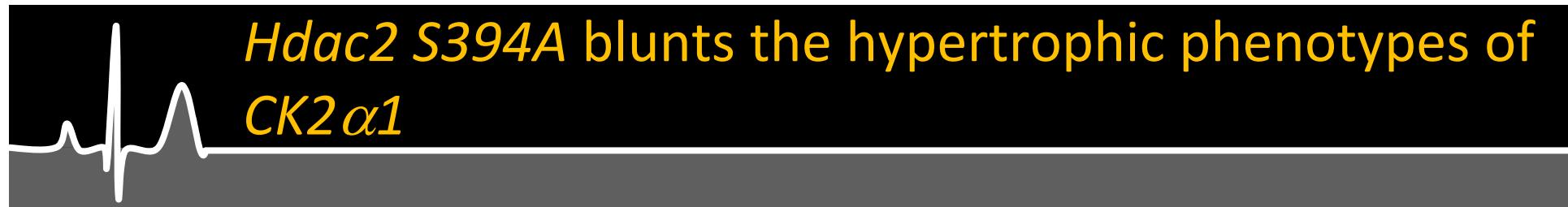
E

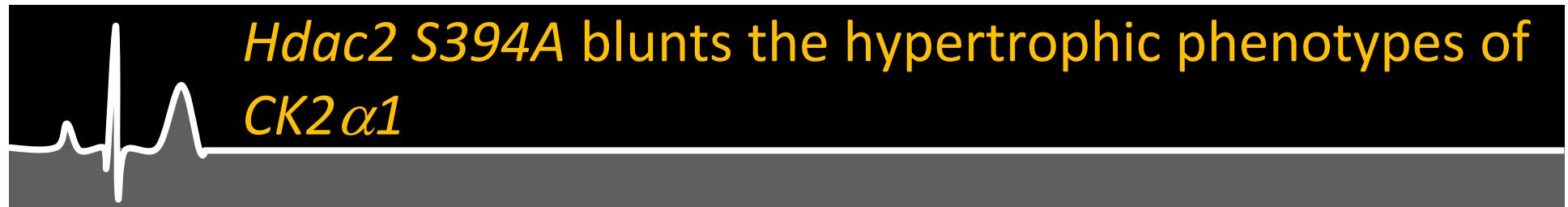


F

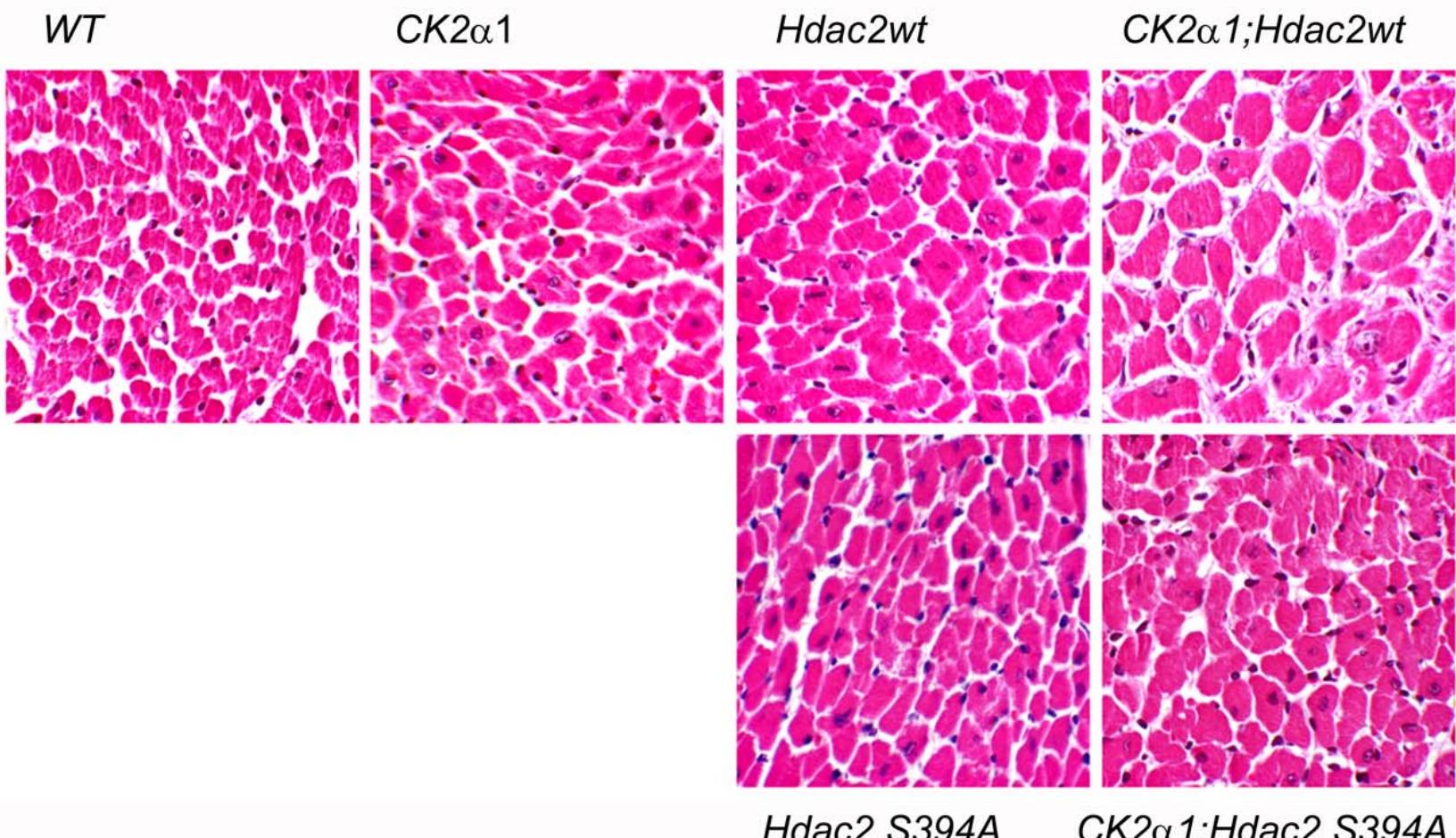








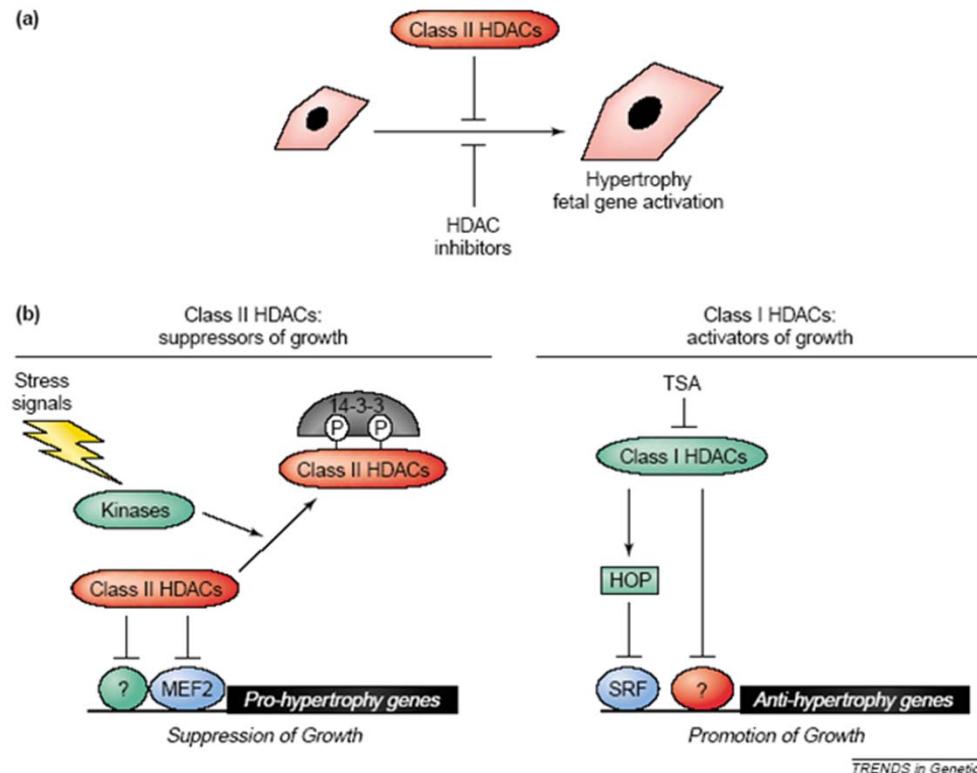
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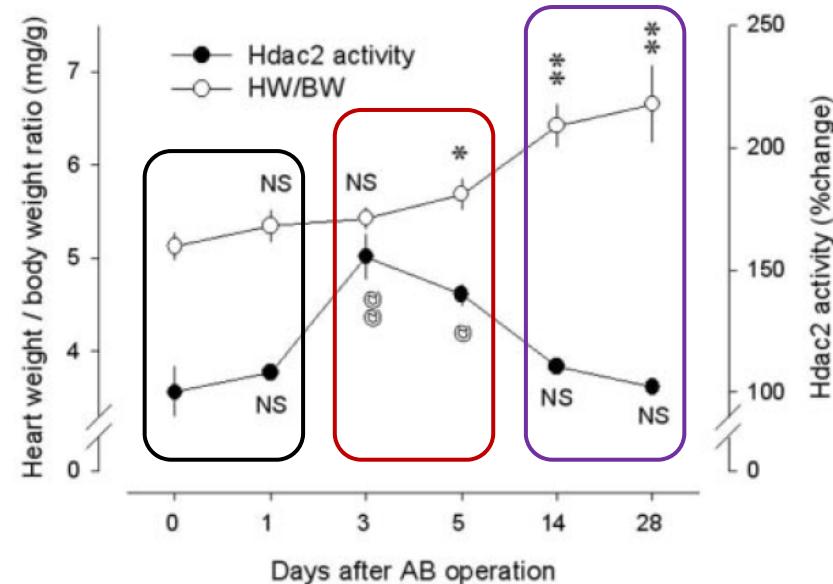
Eom et al., *Circulation* 123(21):2392-2403, 2011



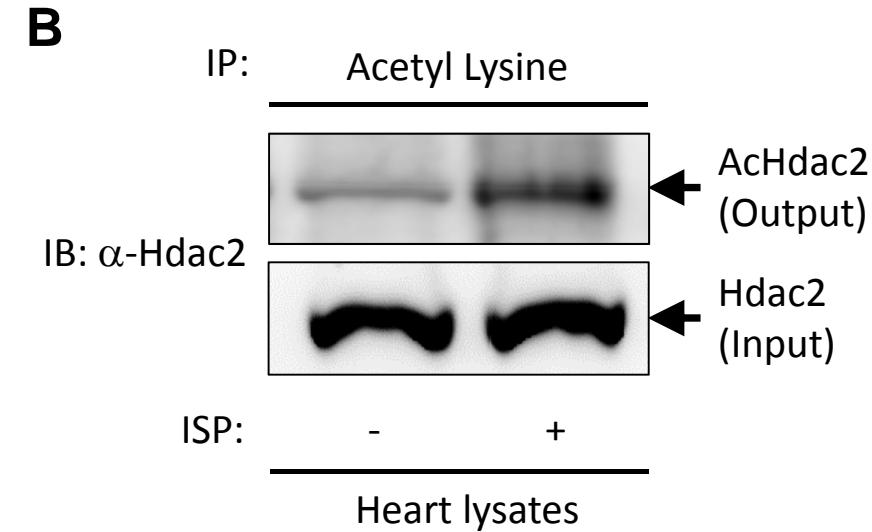
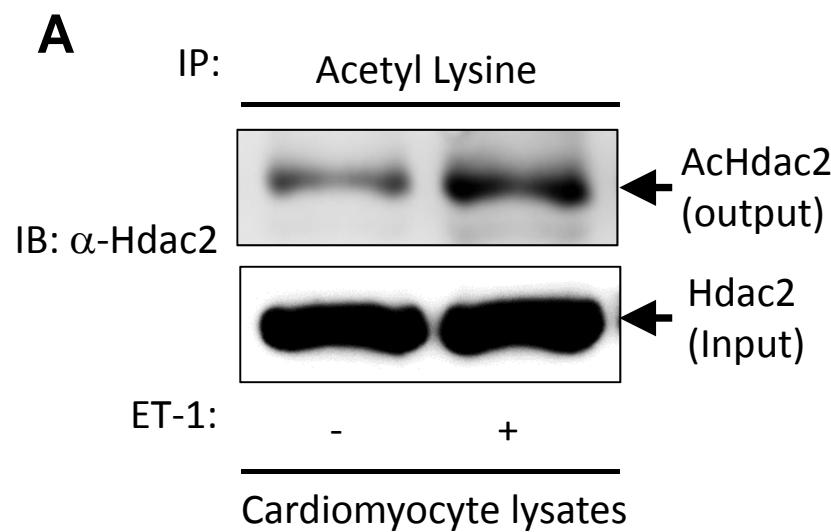
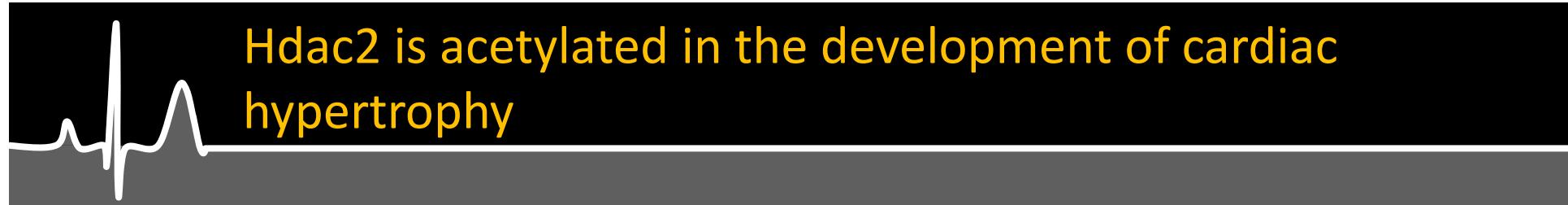
# Crosstalk between Hdac2 and Class IIa HDACs

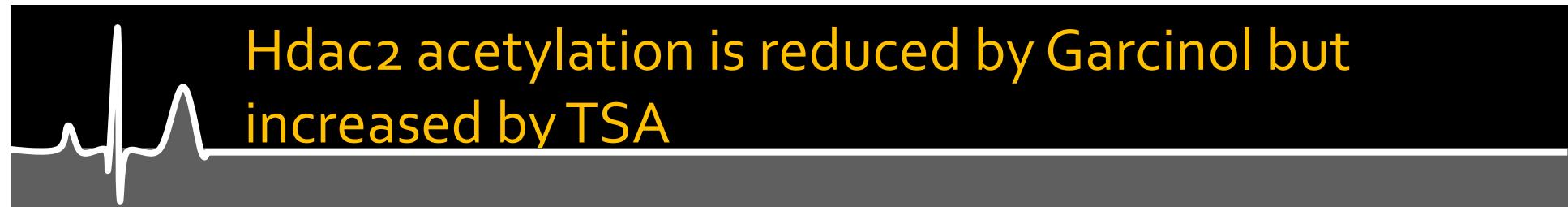


McKinsey TA and Olson EN. *Trends Genet*, 20(4):206-13, 2004

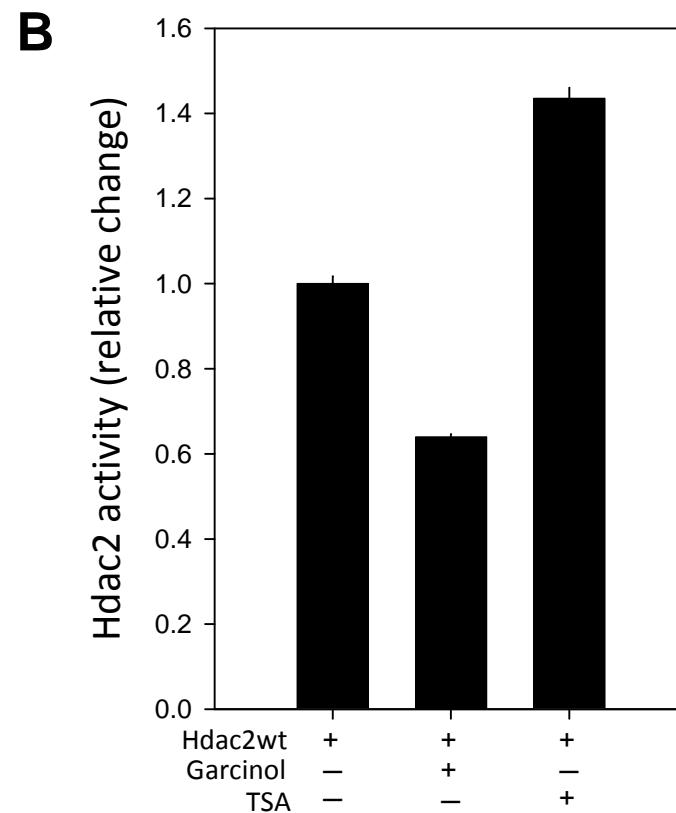
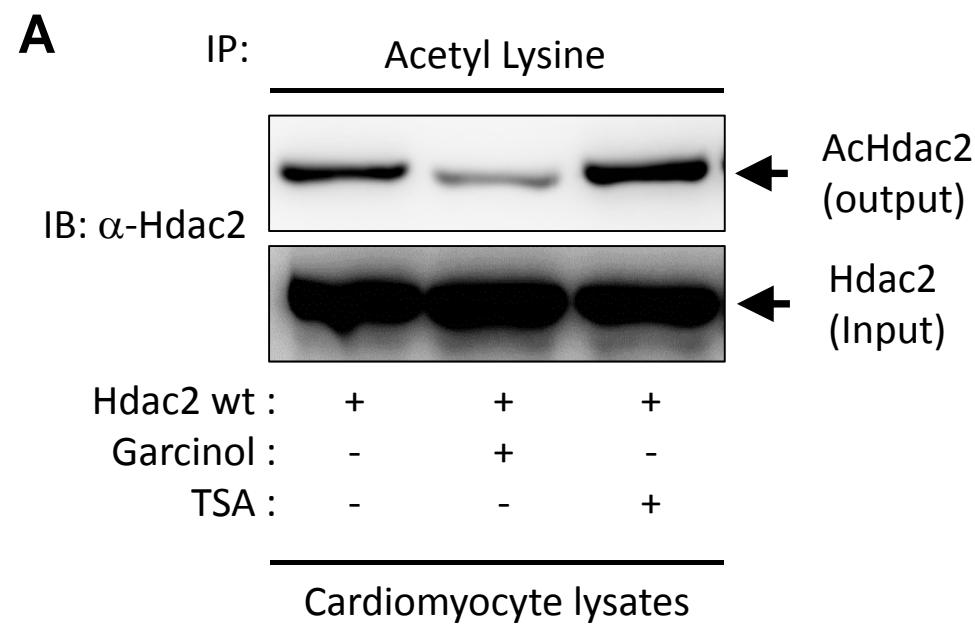


Kee and Eom et al., *Circ Res*, 103(11):1259-69, 2008





# Hdac2 acetylation is reduced by Garcinol but increased by TSA

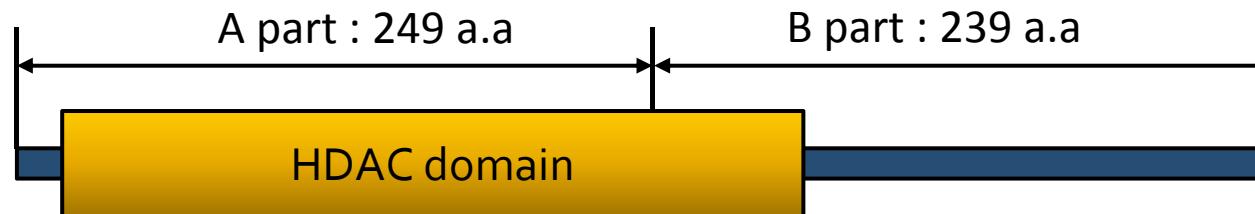


Garcinol : HAT(histone acetyl transferase) inhibitor  
 TSA : HDAC(histone deacetylase) inhibitor

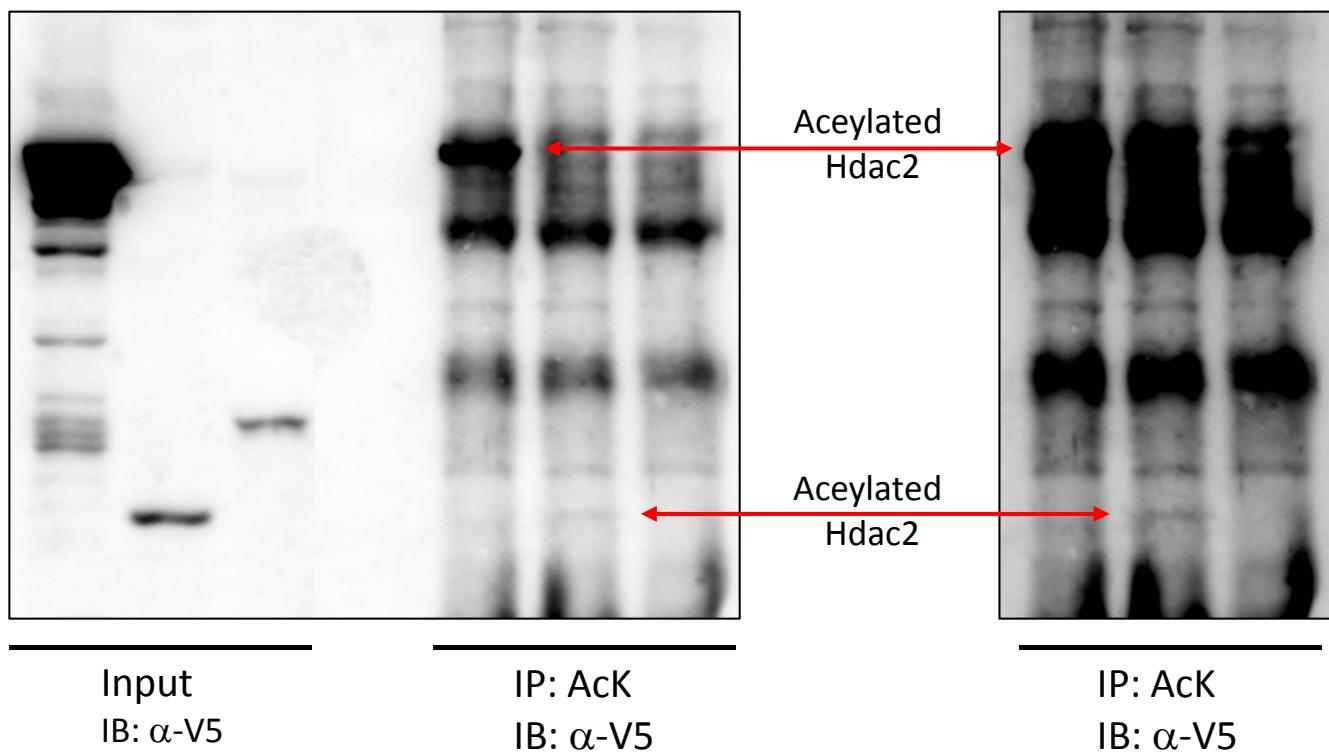
Which residue(s) is(are) the  
acetylation target(s)?

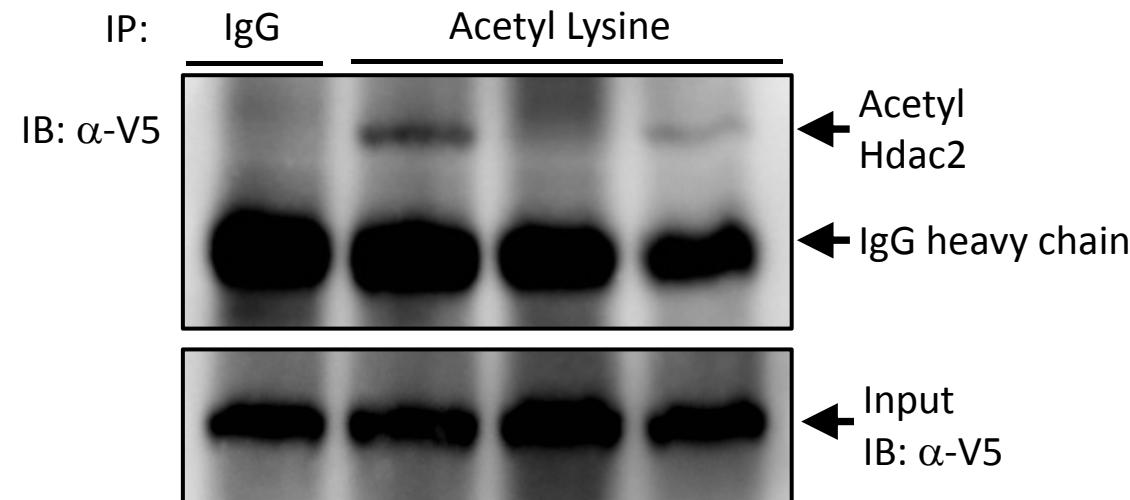


## Acetylation target of Hdac2 is located on the A part



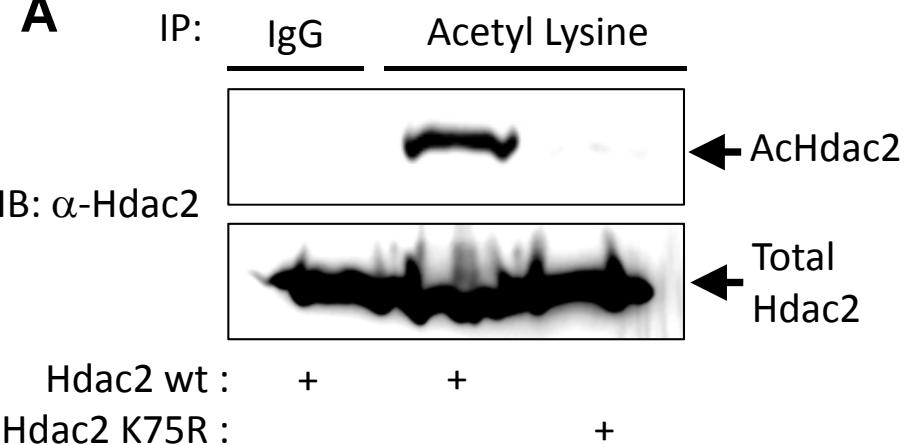
Hdac2 full	+	-	-	+	-	-	+	-	-
Hdac2 A	-	+	-	-	+	-	-	+	-
Hdac2 B	-	-	+	-	-	-	-	-	+



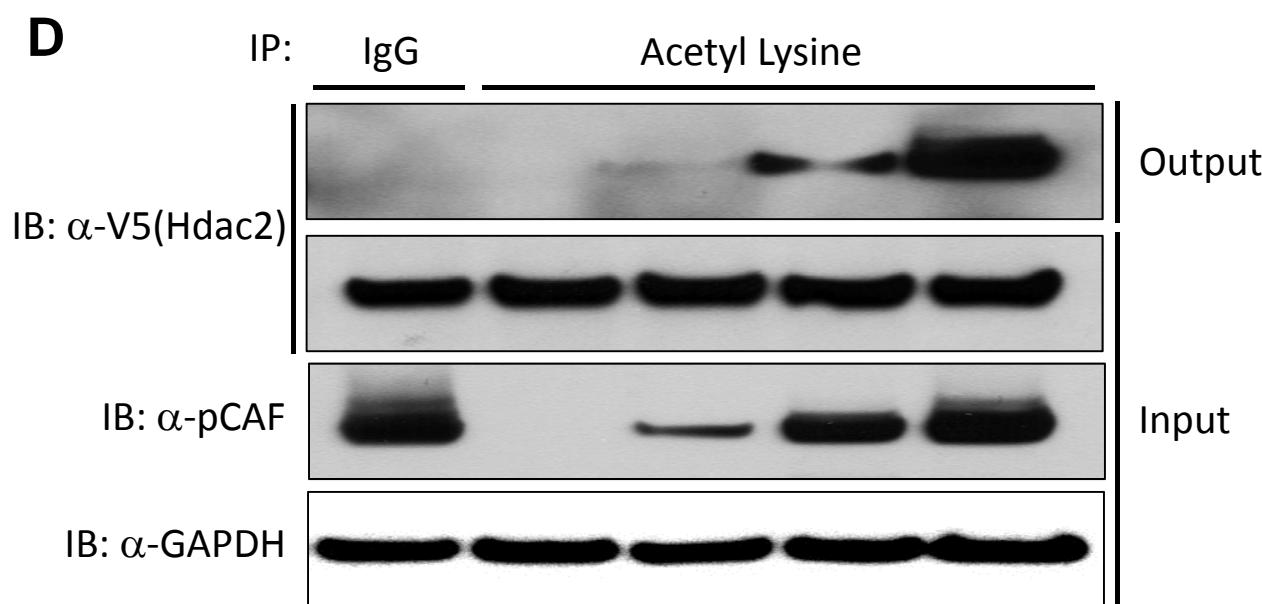
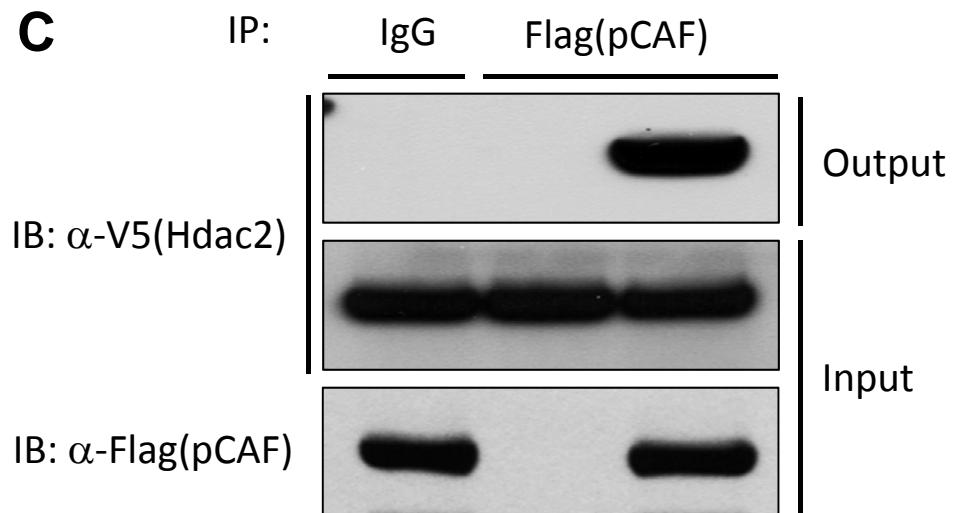


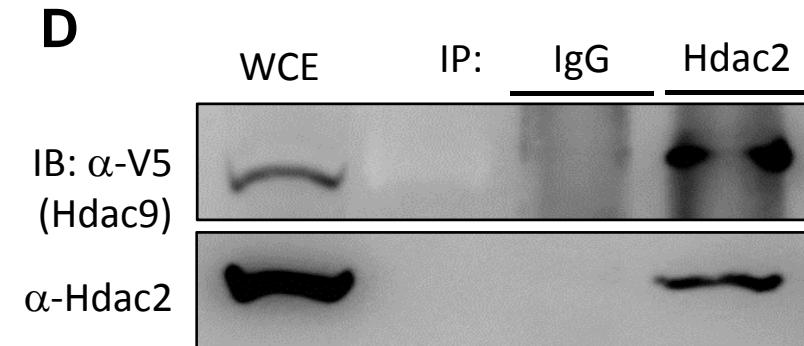
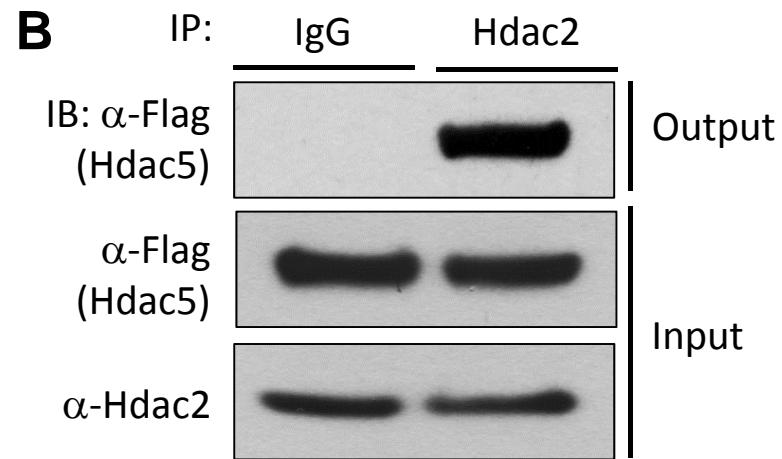
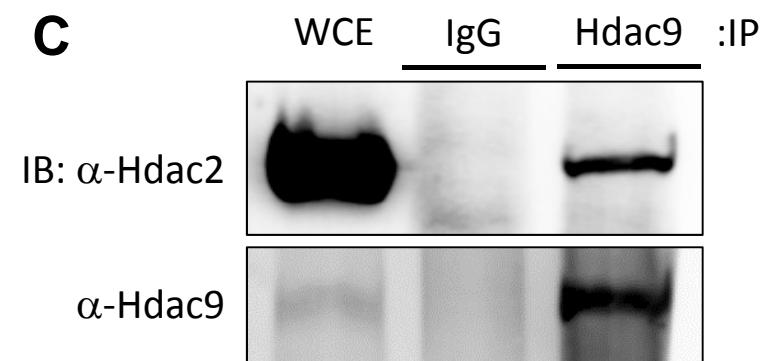
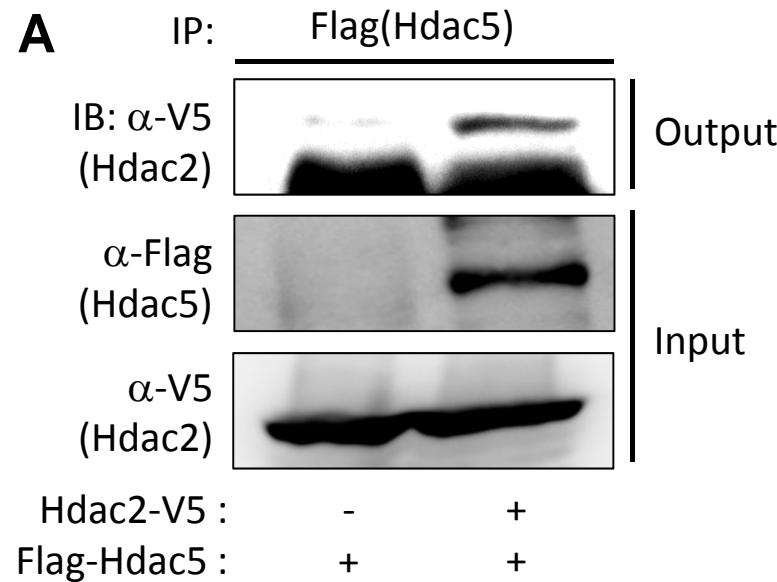
Hdac2 wt :	+	+	-	-
Hdac2 K75R :	-	-	+	-
Hdac2 K90R :	-	-	-	+

A



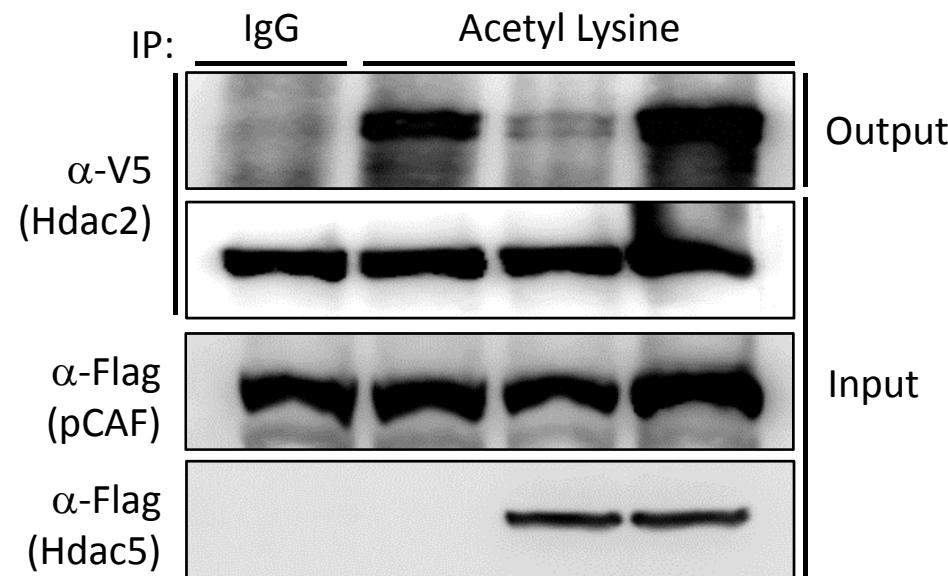
Which HATase?/Which HDAC?







Hdac5 deacetylates Hdac2, which is completely reversed by nonspecific HDAC inhibitor

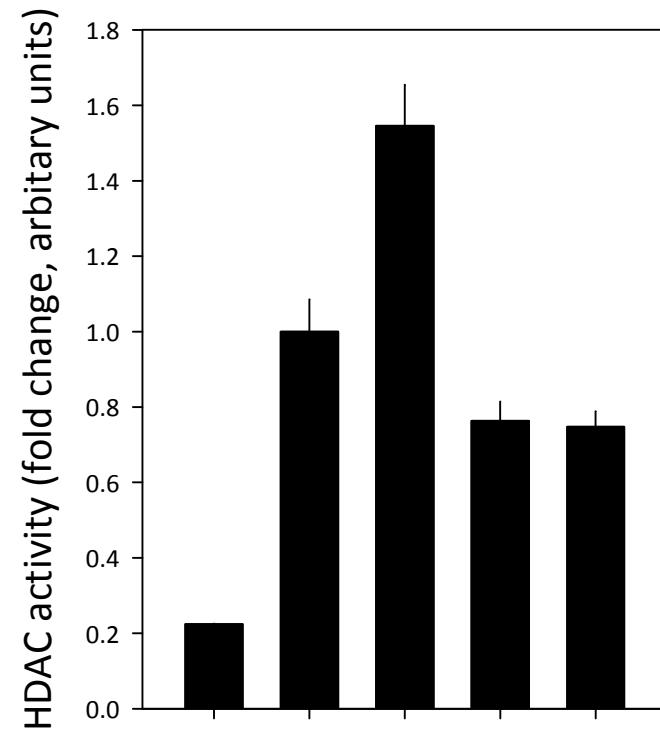


Hdac2-V5	+	+	+	+
Flag-pCAF	+	+	+	+
Hdac5-flag	-	-	+	+
TSA	-	-	-	+

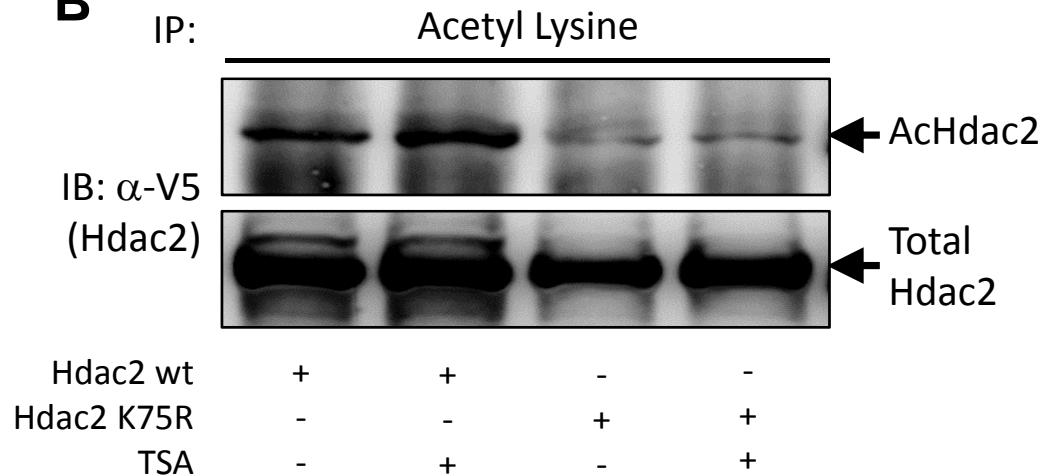
How Acetylation affect to  
enzymatic activity of Hdac2??



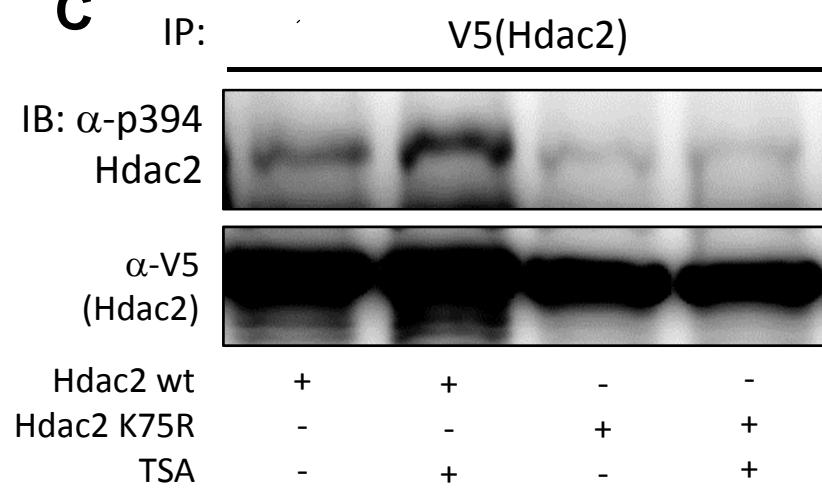
**A**

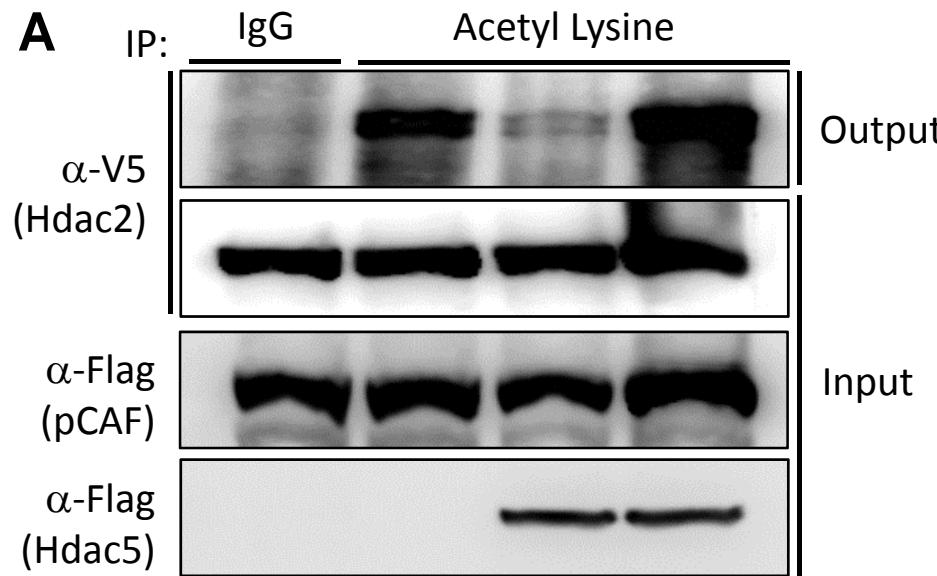


**B**



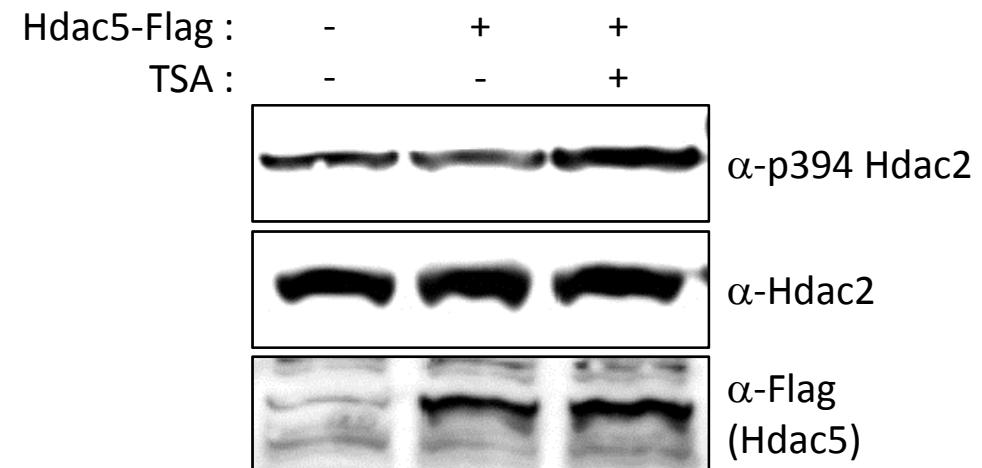
**C**



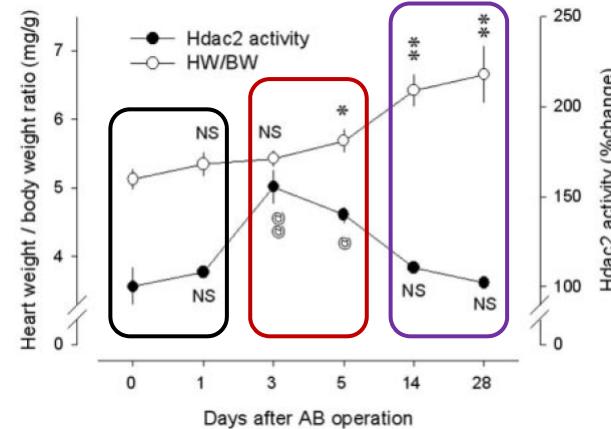


Hdac2-V5	+	+	+	+
Flag-pCAF	+	+	+	+
Hdac5-flag	-	-	+	+
TSA	-	-	-	+

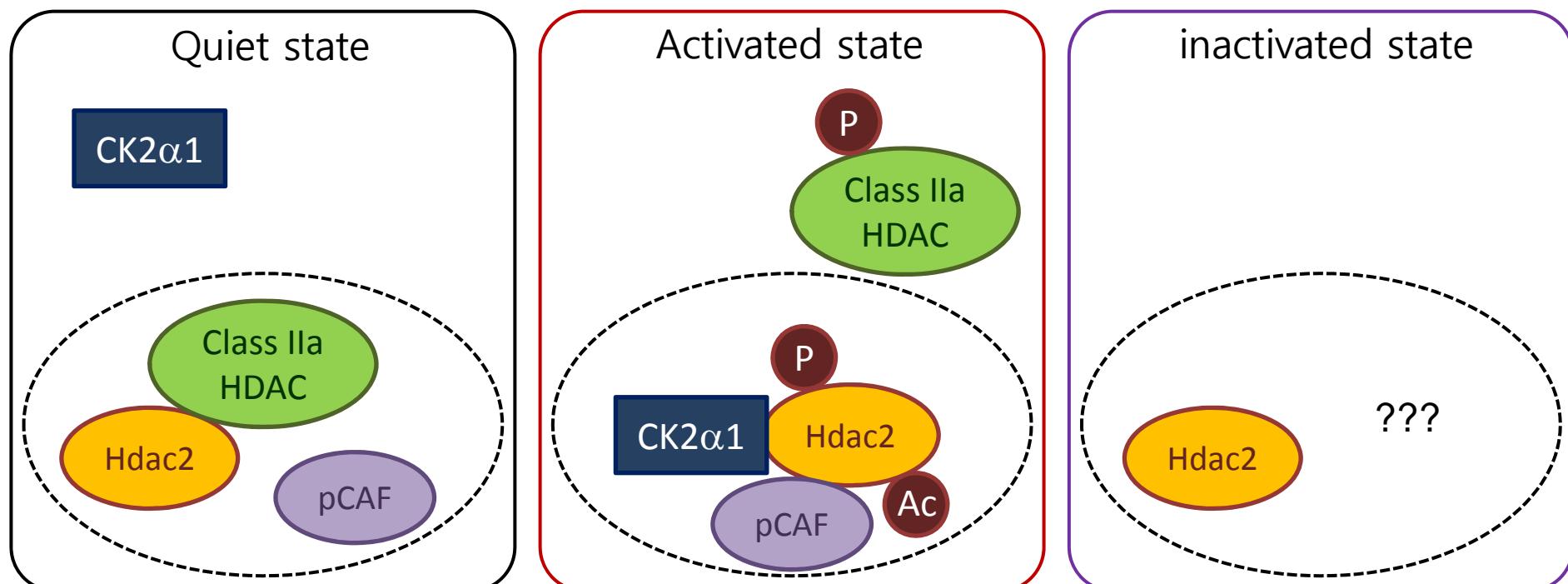
**B**



Eom et al., *in preparation*



Kee and Eom et al., *Circ Res*,  
103(11):1259-69, 2008



Thank you for your attention!!